



Journal Club

Medecine-Science

Session III: Tissue Engineering

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CRI, 10/27/2010



Introduction

- Organ transplantation is often the only treatment for final stage disease
- Shotcomings
 - Availability
 - The need for immunosuppression
- Tissue engineering
- Scaffold based airway and lung tissue engineering



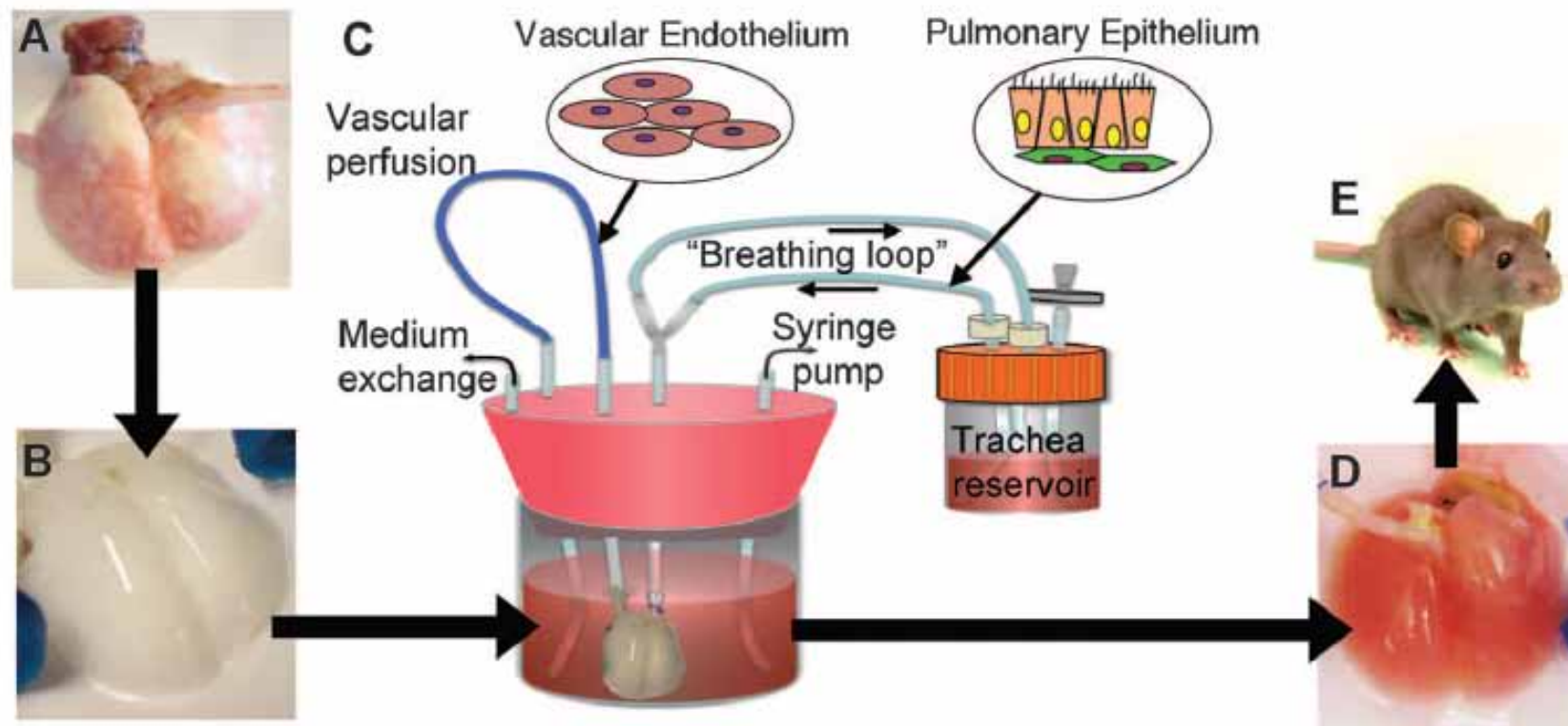
Tissue-Engineered Lungs for in Vivo Implantation

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Tissue-Engineered Lungs for in Vivo Implantation

I Overall Strategy





Tissue-Engineered Lungs for in Vivo Implantation

II Preparation of a Decellularized Lung Scaffold



Tissue-Engineered Lungs for in Vivo Implantation

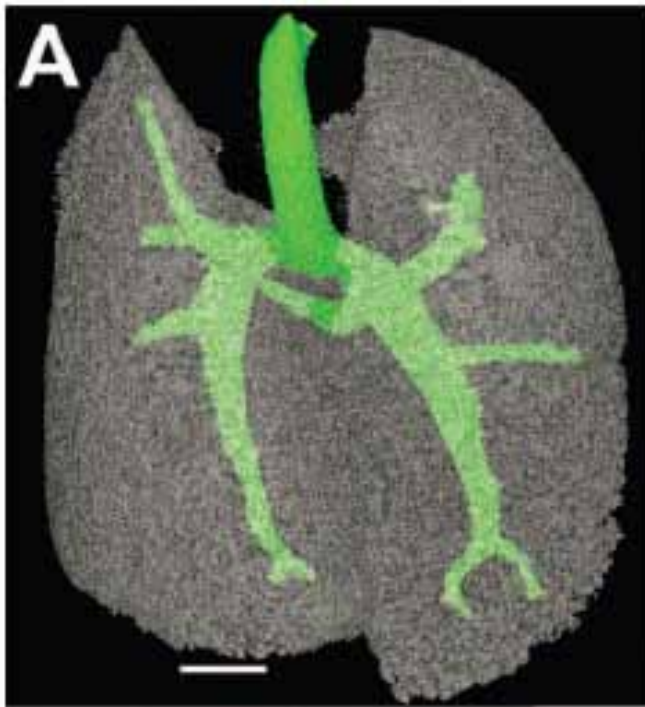
II Preparation of a Decellularized Lung Scaffold

- Lung Tissue was harvested from adult Fischer 344 rats
- The lungs were treated in the vascular and airway compartments with a **detergent solution**
 - 3-[(3-cholamidopropyl)dimethylammonio]-1-propanesulfonate (CHAPS)
 - In a phosphate buffer at 1.0 salt concentration
- Vascular perfusion was maintained < 20mmHg with exposure time 2-3h.

Tissue-Engineered Lungs for in Vivo Implantation

II Preparation of a Decellularized Lung Scaffold

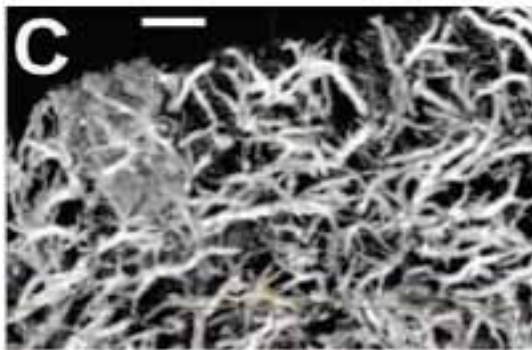
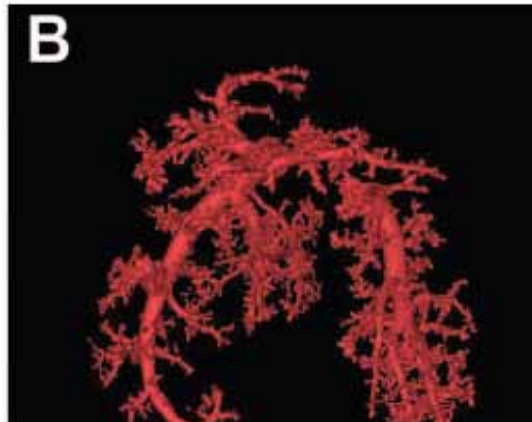
- Intact Lung Architecture



3D-microCT

Tissue-Engineered Lungs for in Vivo Implantation

II Preparation of a Decellularized Lung Scaffold

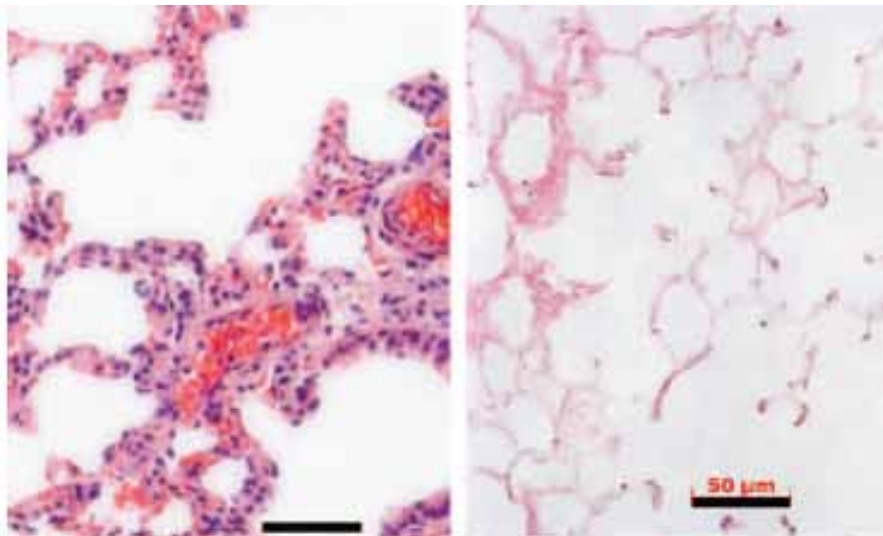


- Intact Lung Architecture
- Intact Arterial tree and microvasculature

3D-microCT angiography

Tissue-Engineered Lungs for in Vivo Implantation

II Preparation of a Decellularized Lung Scaffold



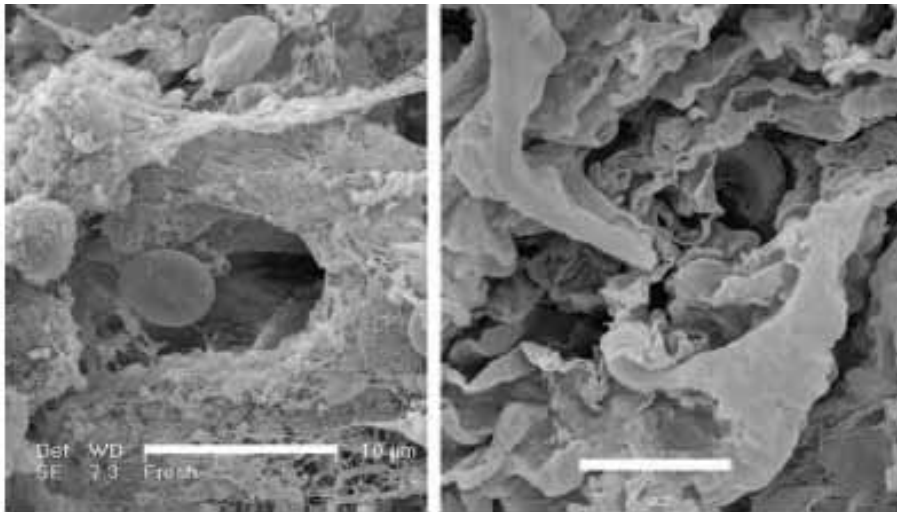
- Intact Lung Architecture
- Intact Arterial tree & microvasculature
- Cells & nuclear material removed
- Alveolas septal architecture conserved

Hematoxylin & Eosin staining

Tissue-Engineered Lungs for in Vivo Implantation

II Preparation of a Decellularized Lung Scaffold

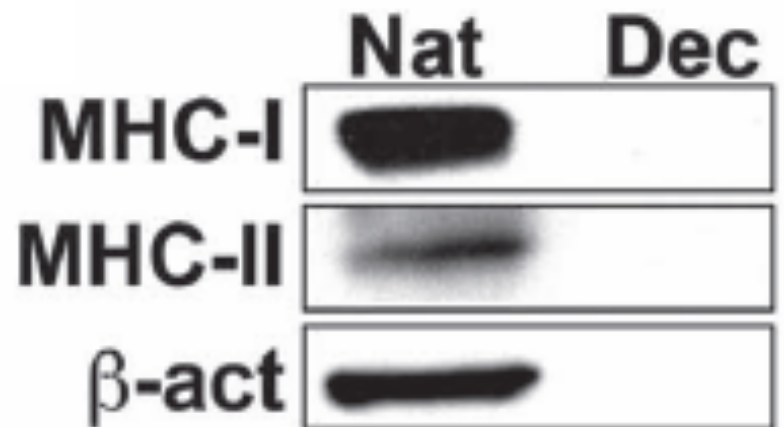
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Scanning Electron Microscopy

Tissue-Engineered Lungs for in Vivo Implantation

II Preparation of a Decellularized Lung Scaffold

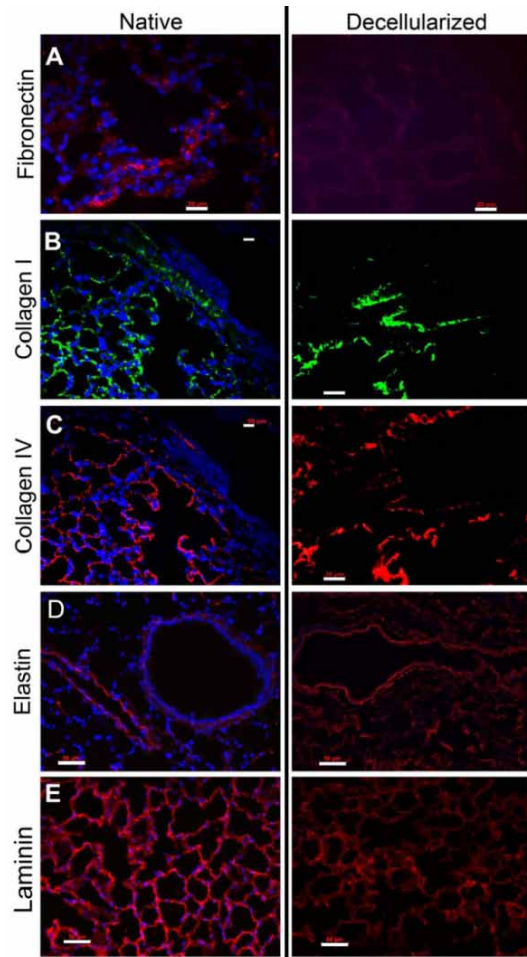


Immunoblotting

- Intact Lung Architecture
- Intact Arterial tree & microvasculature
- Cells & nuclear material removed
- Alveolas septal architecture conserved
- Matrix was depleted of MHC I & II as well as β actin

Tissue-Engineered Lungs for in Vivo Implantation

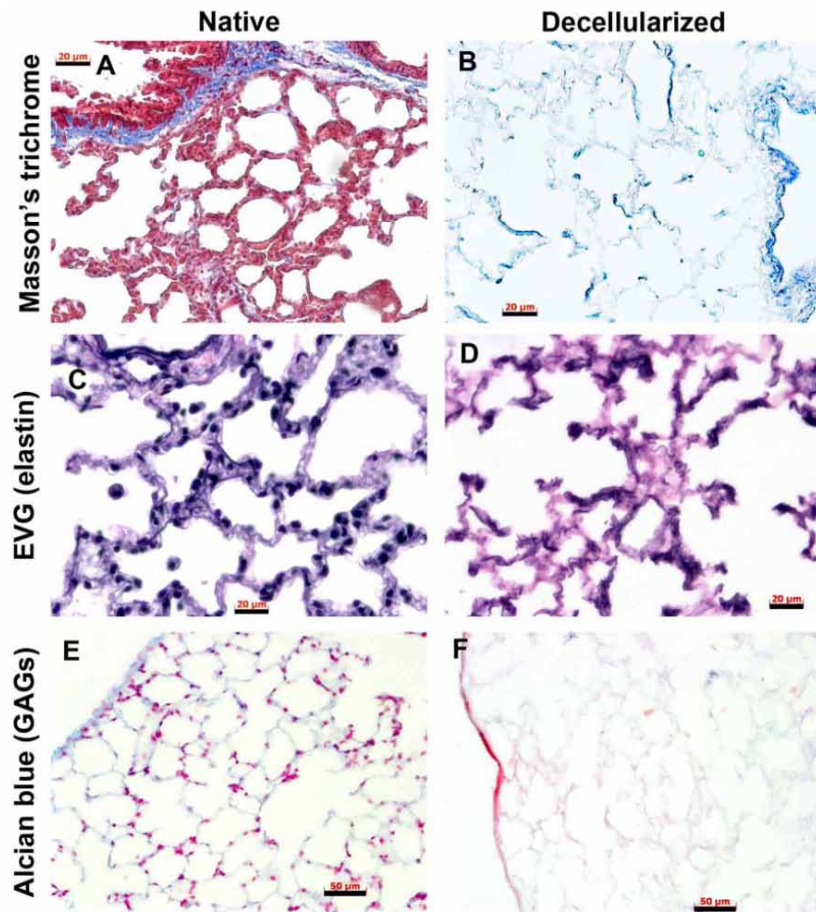
II Preparation of a Decellularized Lung Scaffold



- Intact Lung Architecture
- Intact Arterial tree & microvasculature
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- Matrix was depleted of MHC I & II as well as β actin
- Matrix retains Collagen, Elastin and Laminin

Tissue-Engineered Lungs for in Vivo Implantation

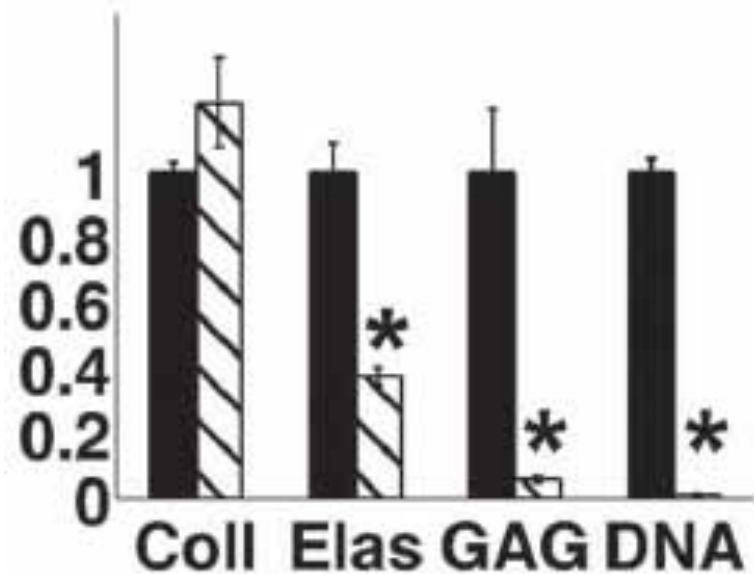
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Tissue-Engineered Lungs for in Vivo Implantation

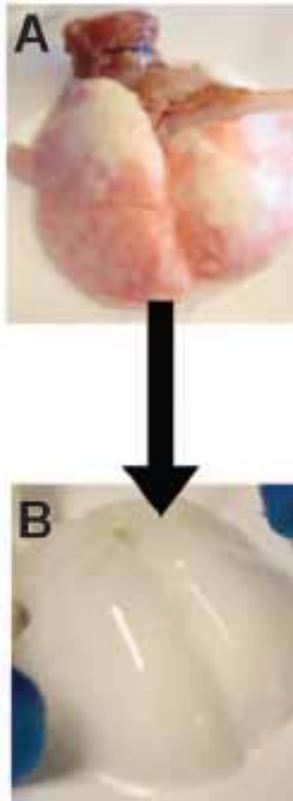
II Preparation of a Decellularized Lung Scaffold



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- Matrix retains Collagen, Elastin and Laminin

Tissue-Engineered Lungs for in Vivo Implantation

II Preparation of a Decellularized Lung Scaffold



Hence, the lung **decellularization** protocol produces an acellular matrix **scaffold** that retains the gross, microstructural, and ultrastructural properties of native lung, yet **removal of antigenic cellular components** is essentially complete

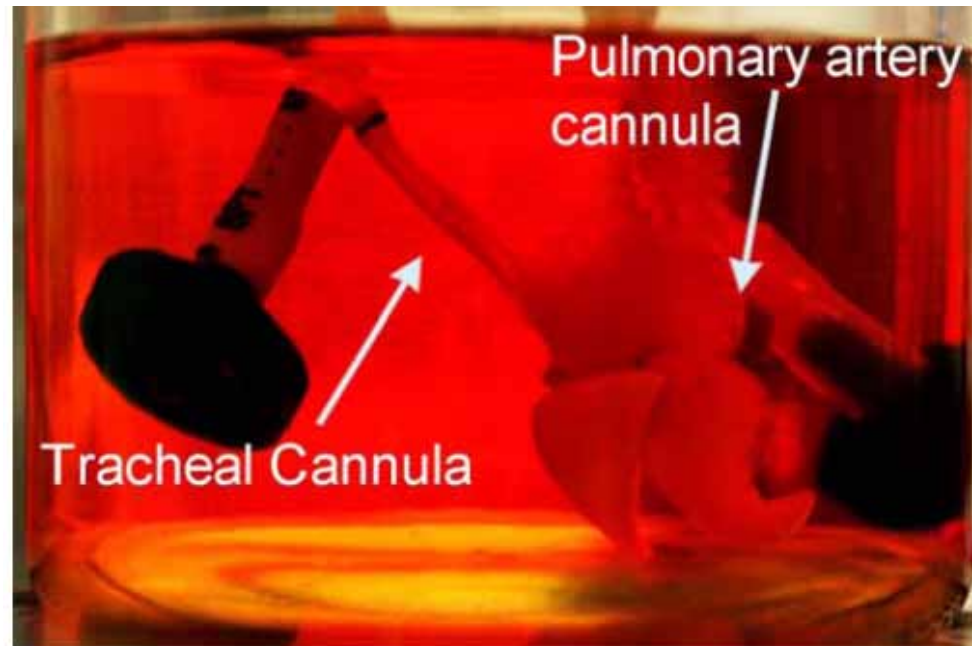


Tissue-Engineered Lungs for in Vivo Implantation

III Properties of the Lung Bioreactor

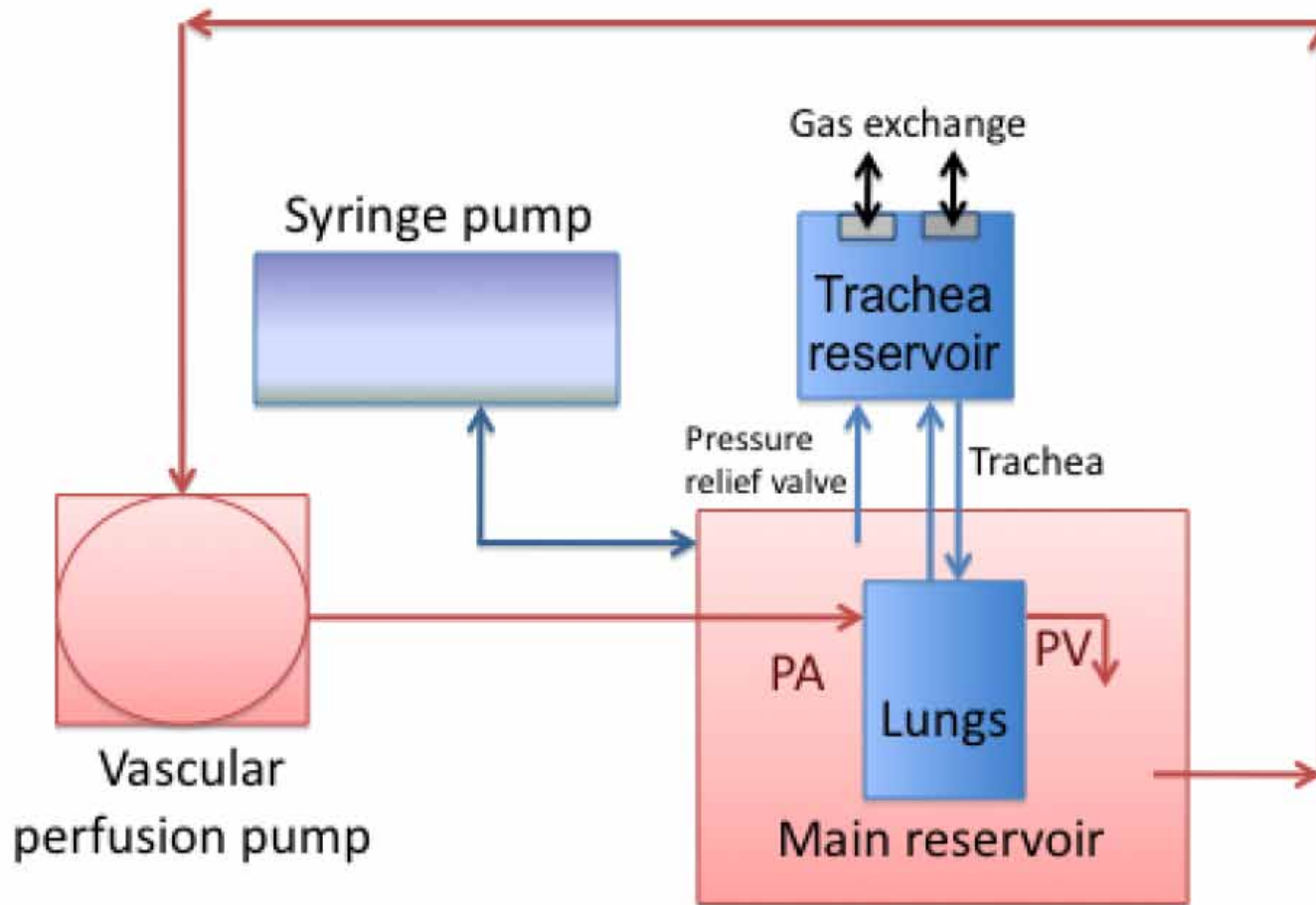
Tissue-Engineered Lungs for in Vivo Implantation

III Properties of the Lung Bioreactor



Tissue-Engineered Lungs for in Vivo Implantation

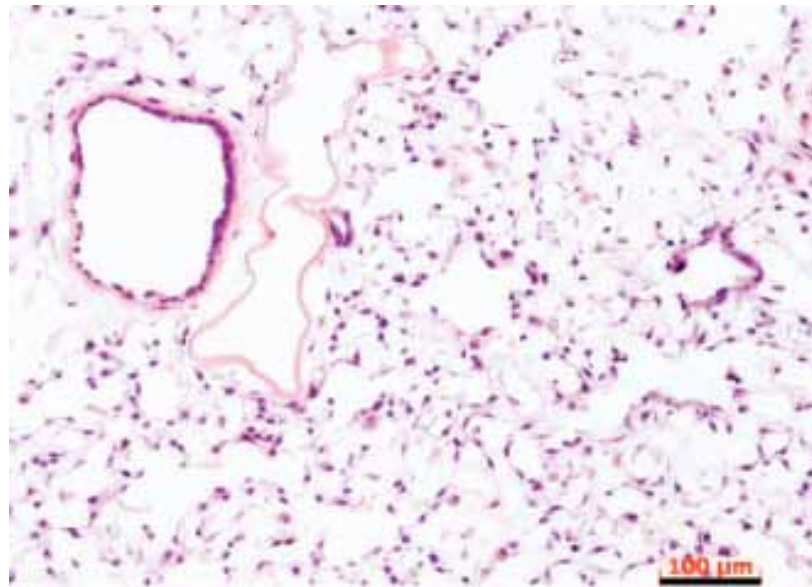
III Properties of the Lung Bioreactor



Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

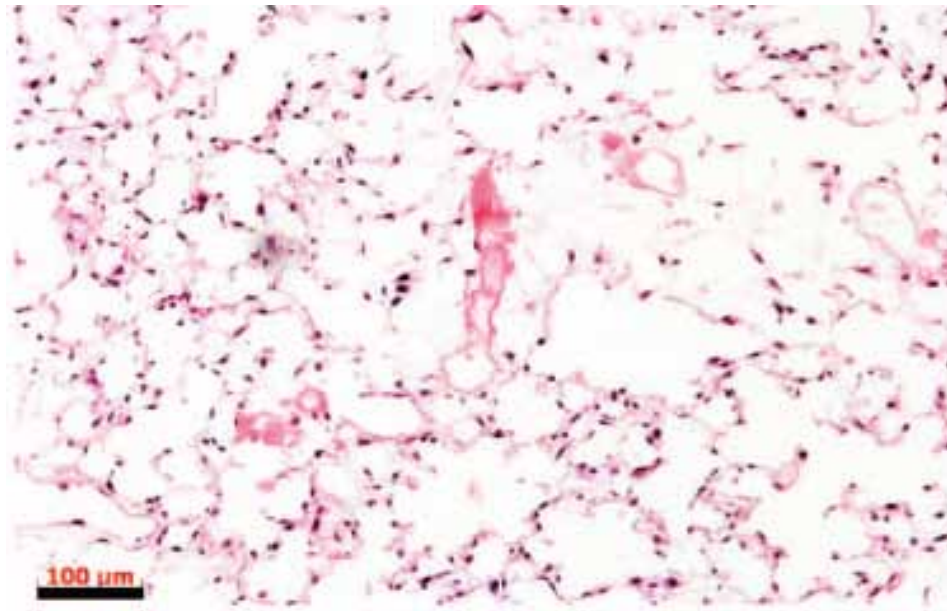
- Lung epithelial cells were isolated from neonatal (<7 day-old) Fischer 344 rats
- The cells were seeded into the airway compartment



Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

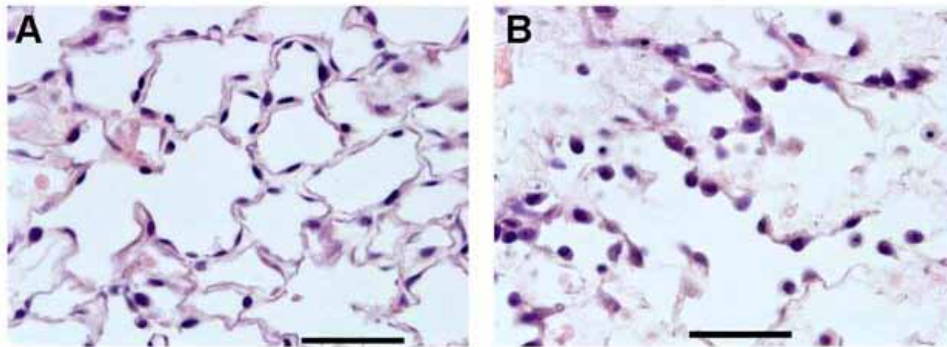
- Rat lung microvascular endothelial cells (RMECs) were seeded into the vascular compartment



Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

- Bioreactor conditions enhanced repopulation

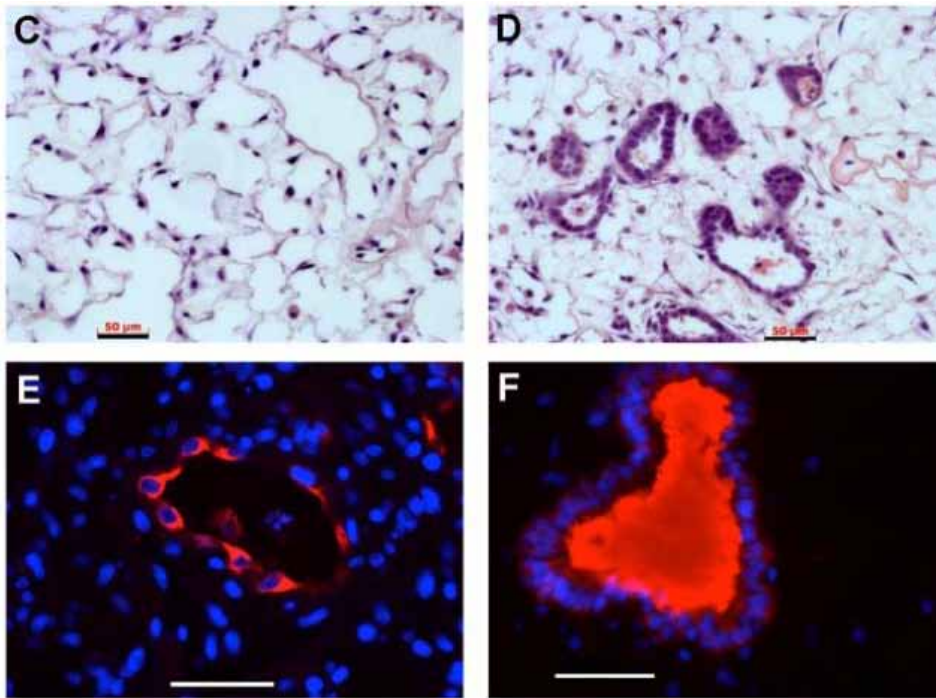


- Vascular perfusion enhanced endothelial adhesion

Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

- Bioreactor conditions enhanced repopulation

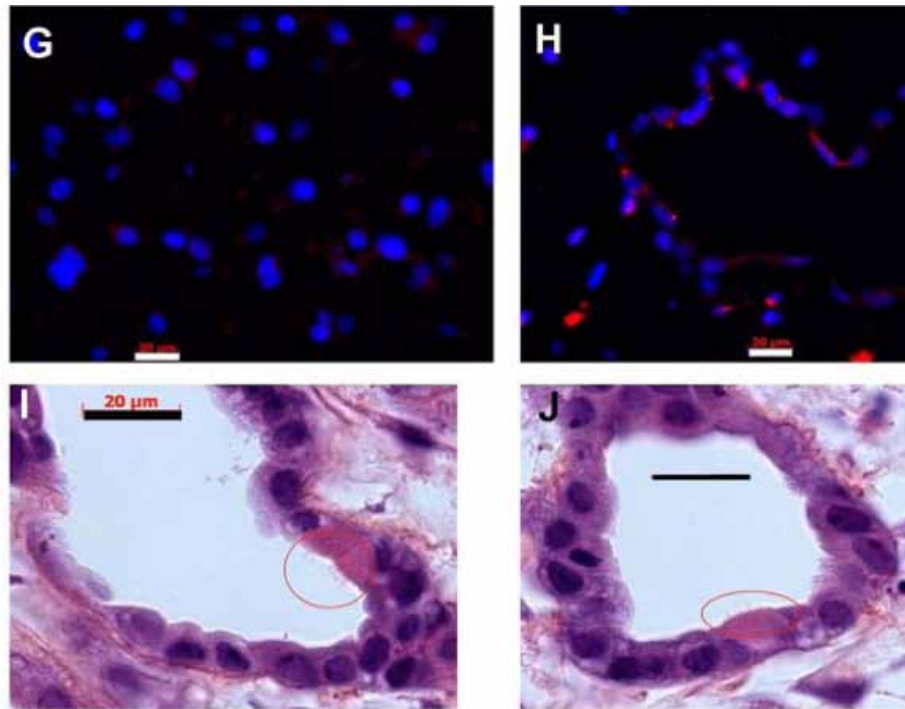


- Vascular perfusion enhanced endothelial adhesion
- Negative pressure ventilation
 - Enhanced adhesion
 - Enhanced clearance

Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

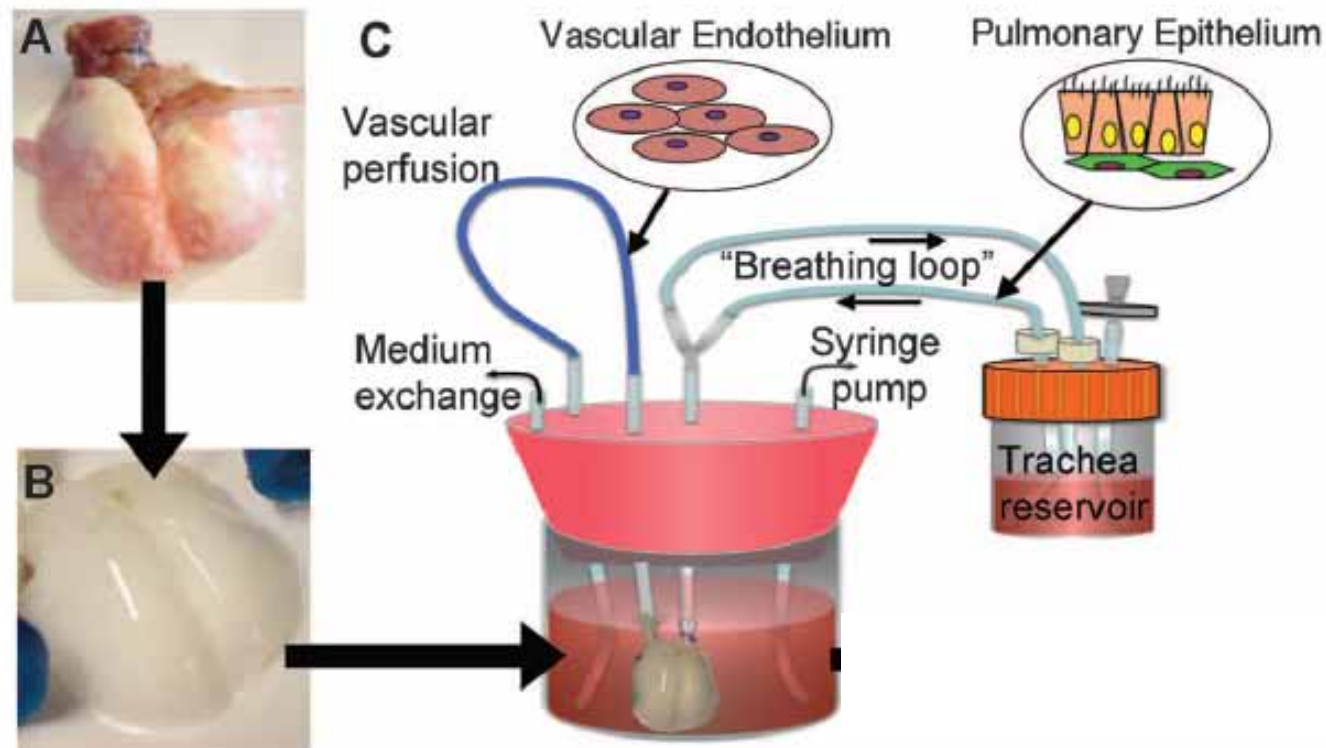
- Bioreactor conditions enhanced repopulation



- Vascular perfusion enhanced endothelial adhesion
- Negative pressure ventilation
- Ventilation with air
 - Type I alveolar epithelial cells
 - Ciliated columnar c.

Tissue-Engineered Lungs for in Vivo Implantation

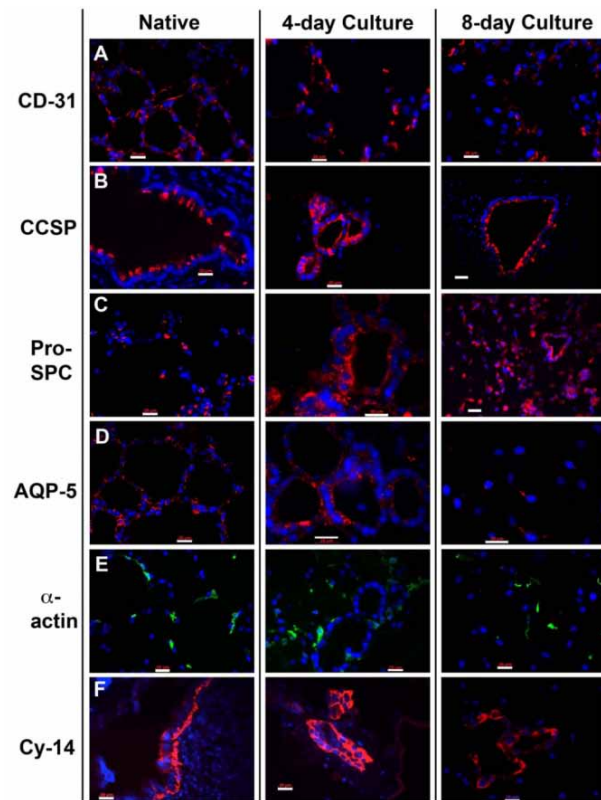
IV Repopulating the acellular matrix



Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

- Characteristics of the Engineered lungs

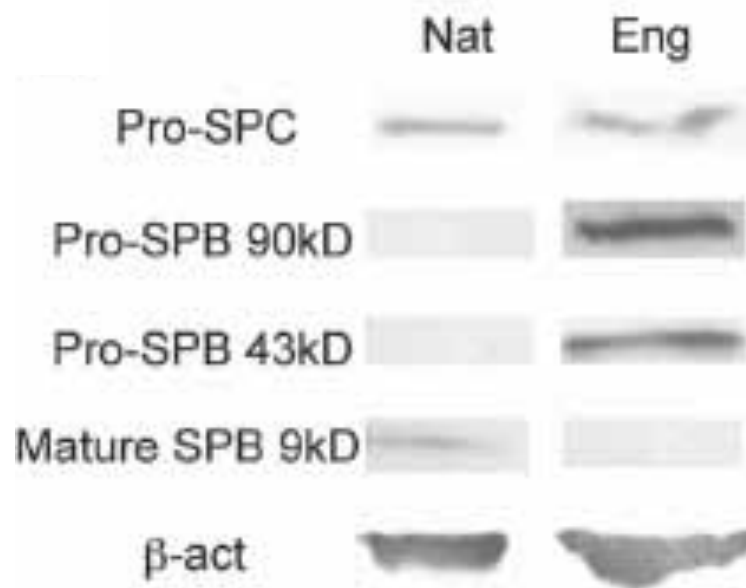


- Cellular composition

Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

- Characteristics of the Engineered lungs

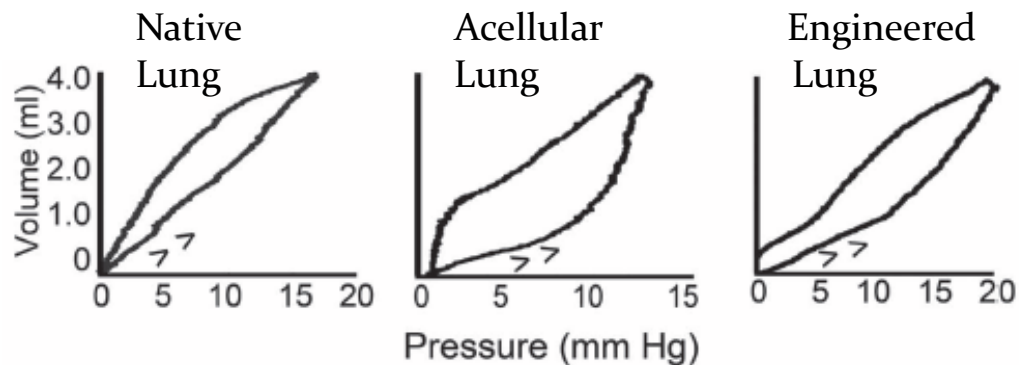


- Cellular composition
- Surfactant secretion

Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

- Characteristics of the Engineered lungs



- Cellular composition
- Surfactant secretion
- Mechanical properties

- Compliance

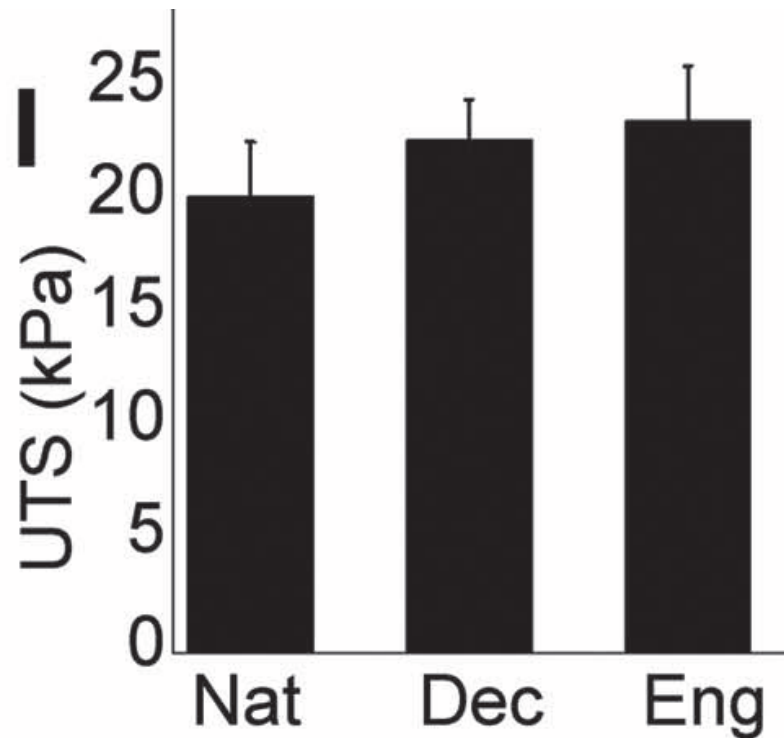
$$C = \frac{\Delta V}{\Delta P}$$

- Native 0.35 ± 0.08 ml/mmHg
- Acellular 0.09 ± 0.02 ml/mmHg
- Engineered 0.14 ± 0.06 ml/mmHg

Tissue-Engineered Lungs for in Vivo Implantation

IV Repopulating the acellular matrix

- Characteristics of the Engineered lungs



- Cellular composition
- Surfactant secretion
- Mechanical properties
 - Compliance
 - Ultimate tensile stress

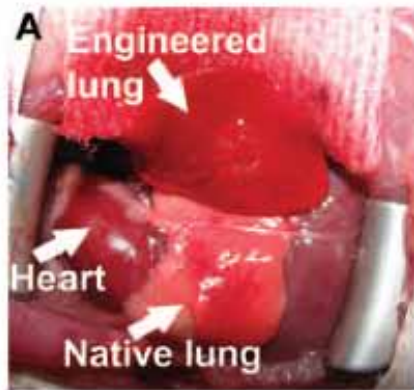


Tissue-Engineered Lungs for in Vivo Implantation

V Implantation of Engineered Lungs into Rats

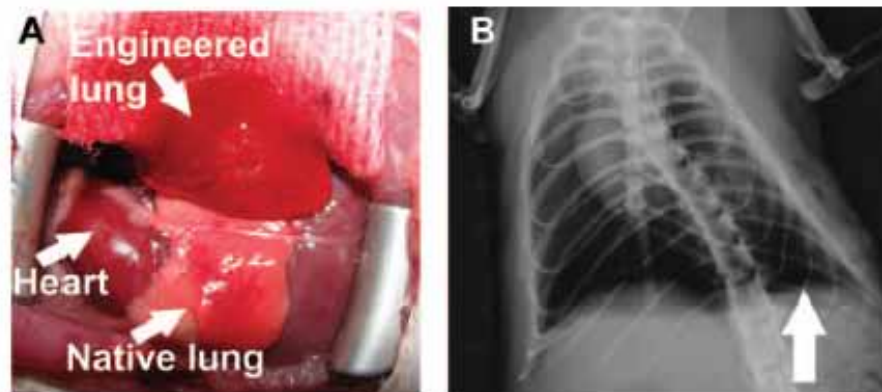
Tissue-Engineered Lungs for in Vivo Implantation

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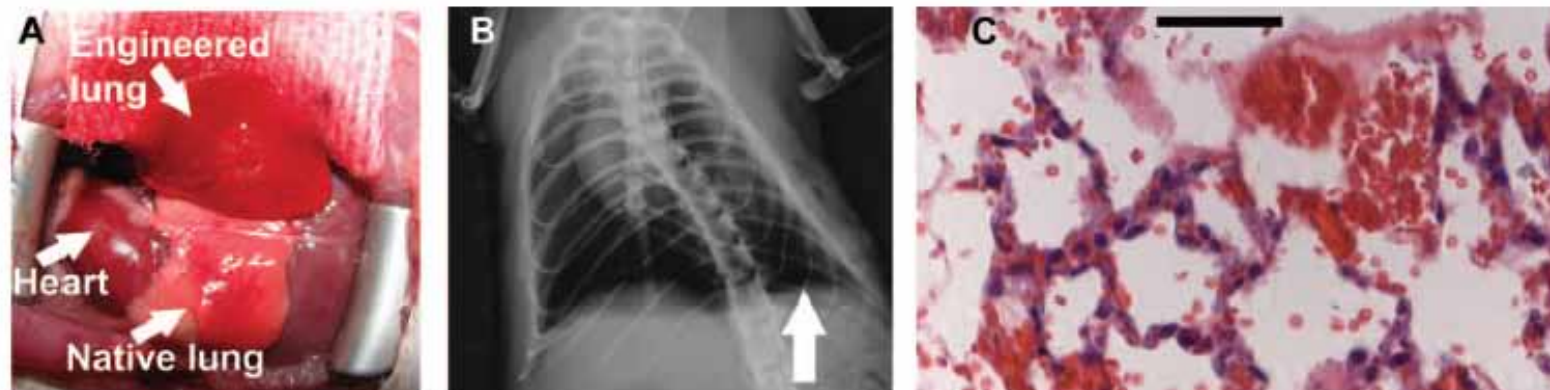
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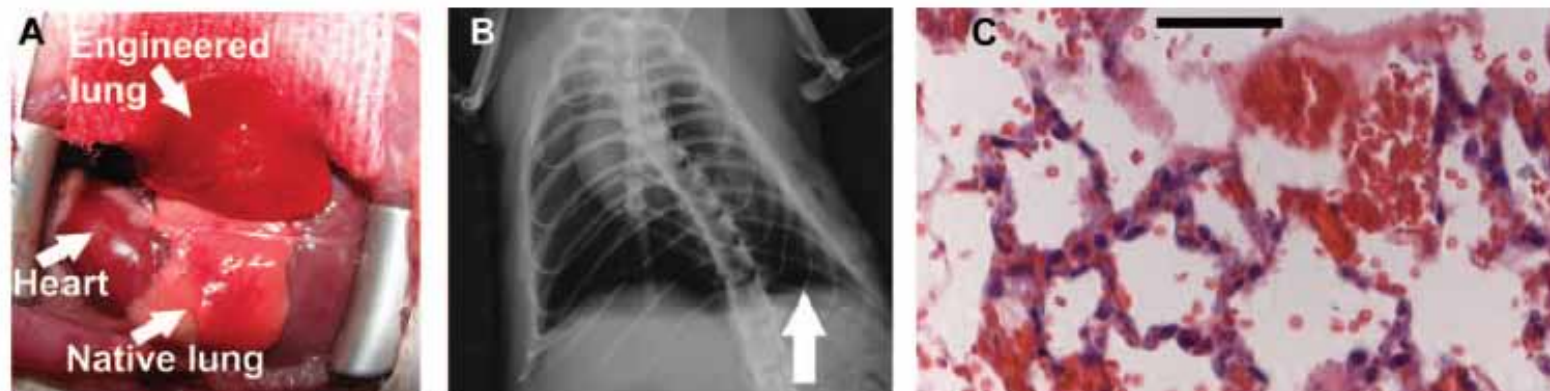
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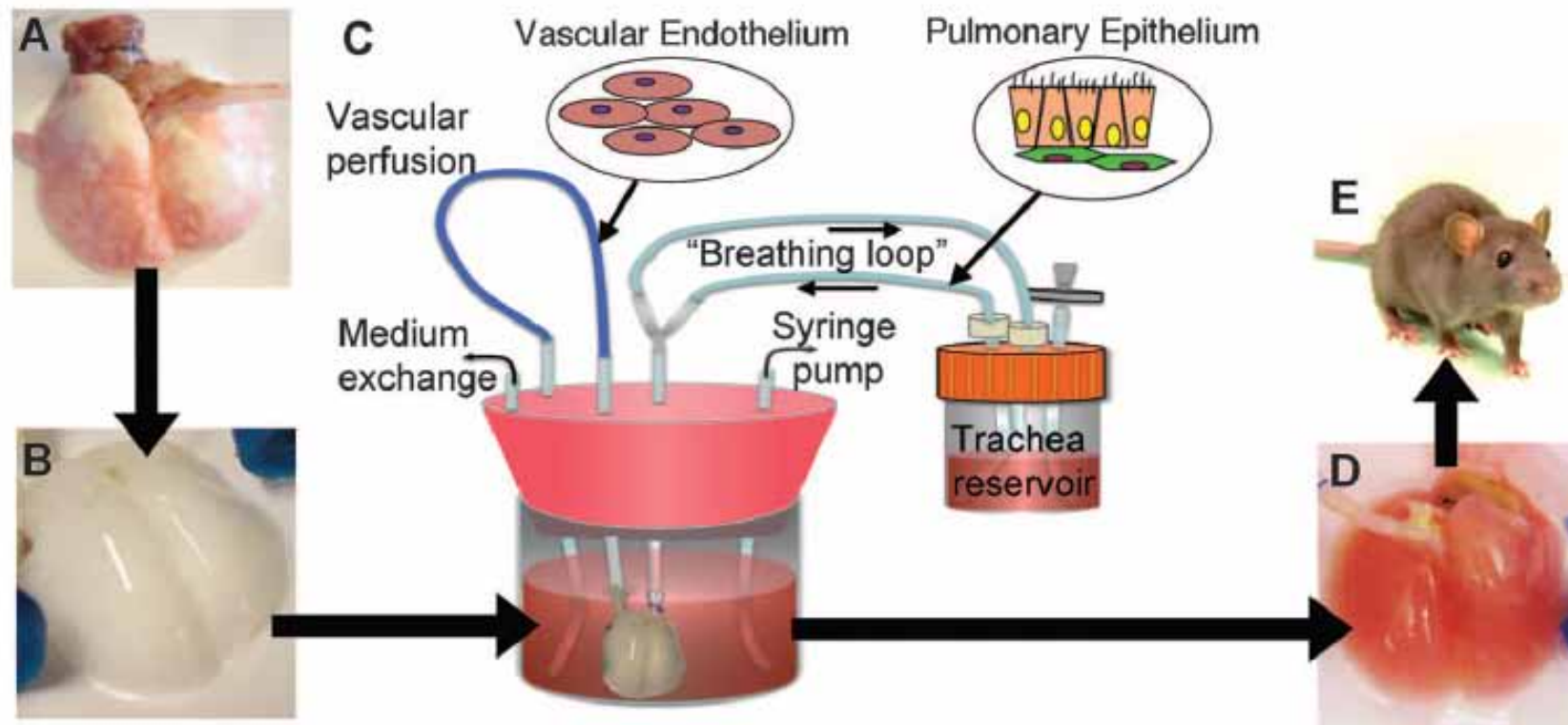
Tissue-Engineered Lungs for in Vivo Implantation

V Implantation of Engineered Lungs into Rats



Sample location	pH	P_{O_2} (mmHg)	O_2 Sat (%)	P_{CO_2} (mmHg)
Pulmonary artery	7.30 ± 0.06	27 ± 7	44 ± 20	41 ± 13
Right pulmonary vein	7.53 ± 0.08	634 ± 69	100 ± 0	20 ± 1
Left (implant) pulmonary vein	7.68 ± 0.28	283 ± 48	100 ± 0	11 ± 5
Mixed pulmonary veins	7.58 ± 0.08	495 ± 174	100 ± 0	18 ± 3

Tissue-Engineered Lungs for in Vivo Implantation






Tissue-Engineered Lungs for in Vivo Implantation

VI Perspectives

- Autologous source of pulmonary epithelium
 - iPSC
 - Resident lung stem cells
- Improved surfactant production for enhanced compliance
- Enhanced alveolar barrier function
- Enhanced vascular endothelial coverage



Clinical transplantation of a tissue-engineered airway

Paolo Macchiarini, Philipp Jungebluth, Tetsuhiko Go, M Adelaide Asnaghi, Louisa E Rees, Tristan A Cogan, Amanda Dodson, Jaume Martorell, Silvia Bellini, Pier Paolo Parnigotto, Sally C Dickinson, Anthony P Hollander, Sara Mantero, Maria Teresa Conconi, Martin A Birchall

Vol 372 Decembre 13 2008. The Lancet



Clinical transplantation of a tissue-engineered airway

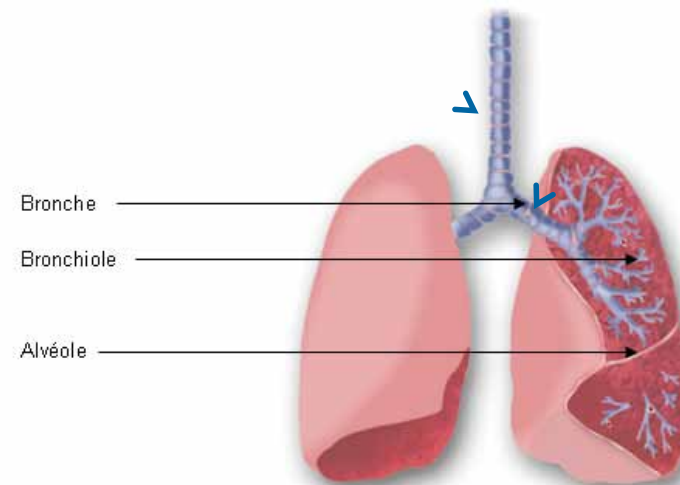
I Introduction

- Large airway defects are a challenge to clinicians because of the lack of efficient methods of treatment
 - Tracheal resection with primary repair is the only curative treatment
 - Resecable length < 30% in children & <6 cm in adults
 - Previous attempts of autologous or synthetic grafts were disappointing
- The aim of this study is to assess the use of a reseeded decellularized tracheal scaffold for the treatment of end-stage airway disease

Clinical transplantation of a tissue-engineered airway

II Medical History

- A 30 year old woman presented in 2004
 - Cough and dysphonia
 - Due to tuberculous infiltration of cervical trachea and entire left main bronchus





Clinical transplantation of a tissue-engineered airway

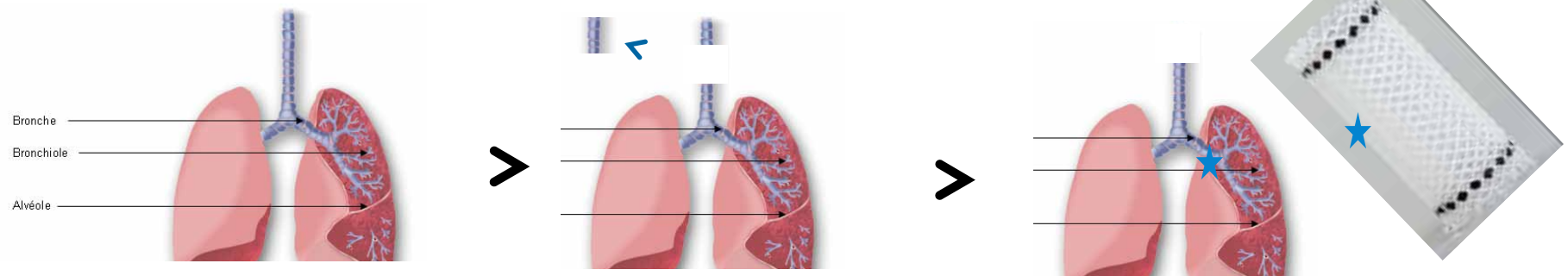
II Medical History

- A 30 year old woman presented in 2004
 - Cough and dysphonia
 - Due to tuberculous infiltration of cervical trachea and entire left main bronchus
 - A CT scan showed a circumferential, near-occlusive 3 cm airway stenosis starting 2cm subglottically and a hypoplastic left main bronchus with expiratory collapse
 - The infection was successfully treated in 6 months

Clinical transplantation of a tissue-engineered airway

II Medical History

- Severe dyspnea persisted
 - Histology: squamous metaplasia without residual infection
 - Diag: Post-tuberculosis chronic tracheitis and secondary severe bronchomalacia of left main bronchus
 - TT: Subglottic tracheal resection/end-to-end anastomosis + Stent in the left main bronchus





Clinical transplantation of a tissue-engineered airway

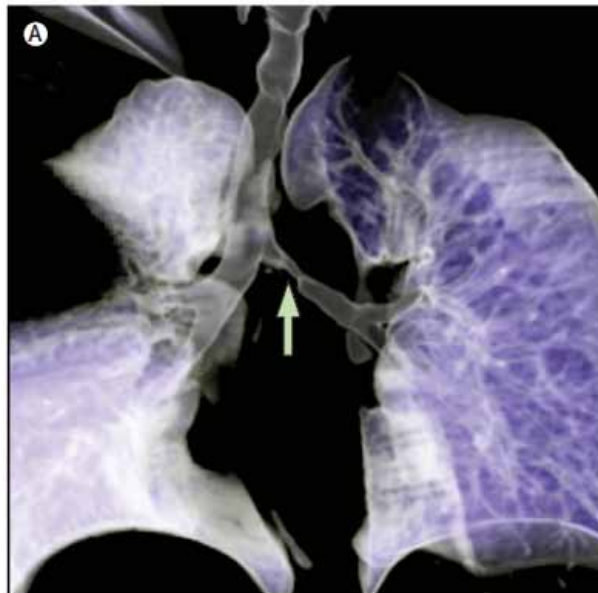
II Medical History

- The stent was poorly tolerated
 - Multiple endoscopic toilets, repositioning and replacement procedures
 - Recurrent episodes of pneumonitis in the left lower lobe
 - Untreatable cough and mucus retention
 - The stent was removed

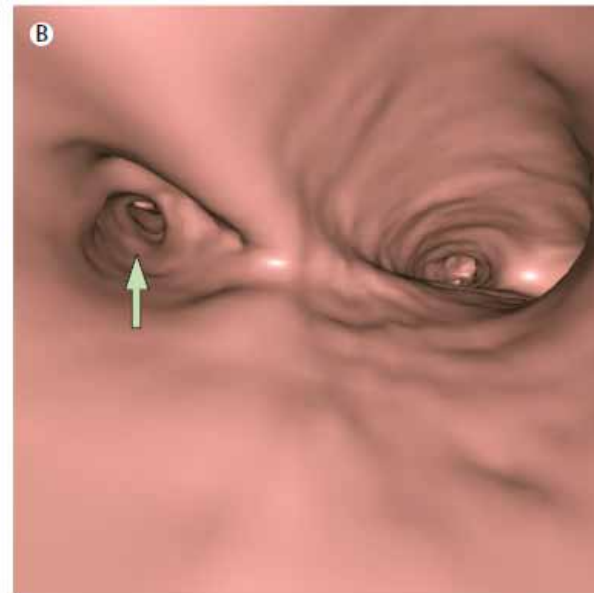
Clinical transplantation of a tissue-engineered airway

II Medical History

- In march 2008, the patient, age 34, was admitted for severe dyspnea (stage III of NYHA classification)



Volume rendering CT-Scan

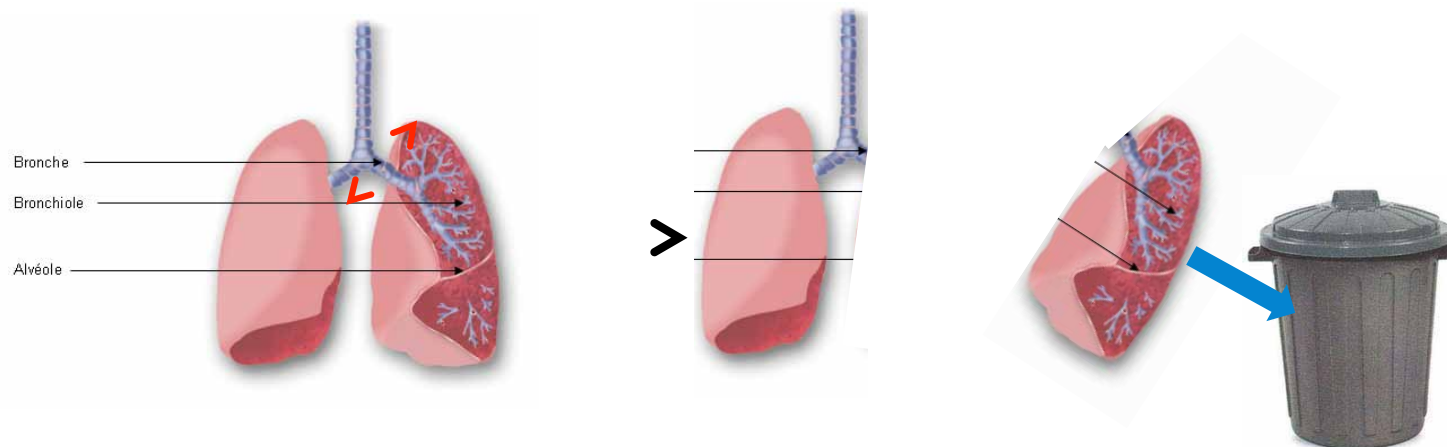


Virtual bronchoscopy

Clinical transplantation of a tissue-engineered airway

II Medical History

- In march 2008, the patient, age 34, was admitted for severe dyspnea (stage III of NYHA classification)
- The only remaining conventional option was
 - left carinal total pneumonectomy





Clinical transplantation of a tissue-engineered airway

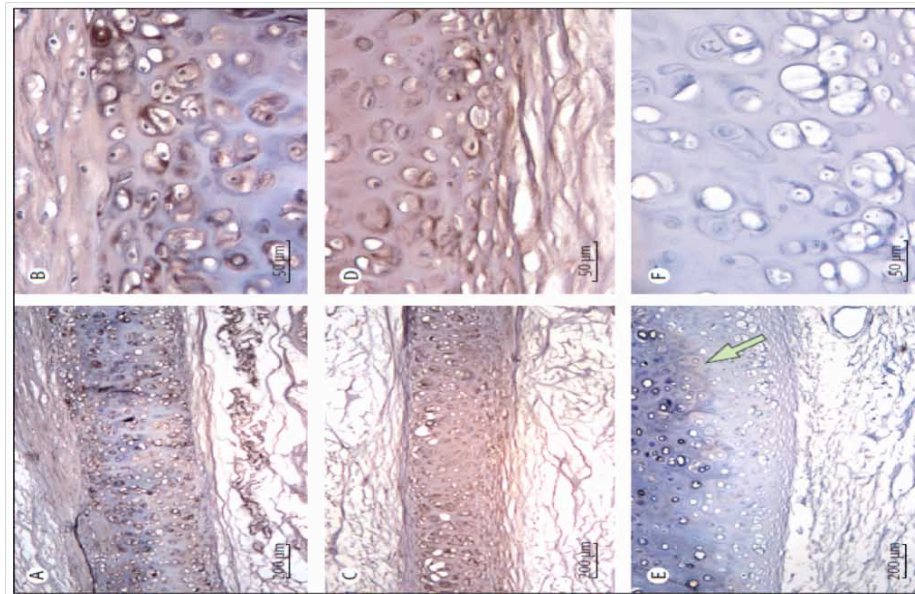
III Strategy

- The authors proposed left main bronchus resection and replacement with a bioengineered human trachea

Clinical transplantation of a tissue-engineered airway

IV Graft Preparation

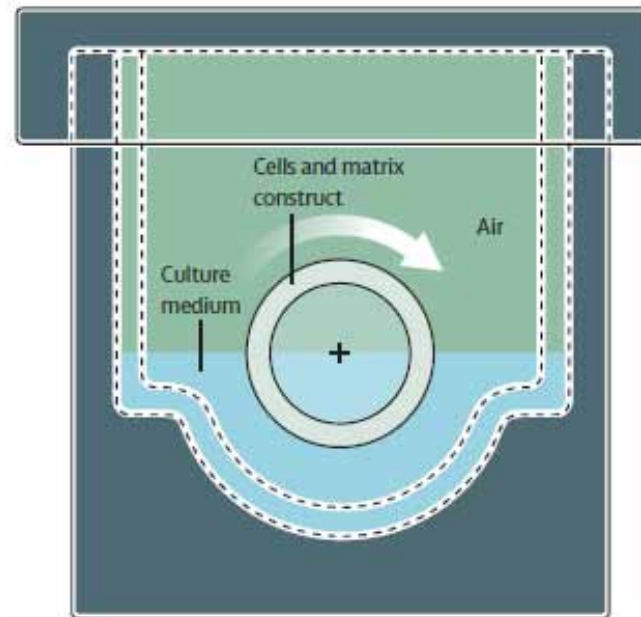
- Preparation of airway matrix
 - 7 cm tracheal segment
 - 25 cycles of decellularization protocol



Clinical transplantation of a tissue-engineered airway

IV Graft Preparation

- Preparation of airway matrix
- Bioreactor design





Clinical transplantation of a tissue-engineered airway

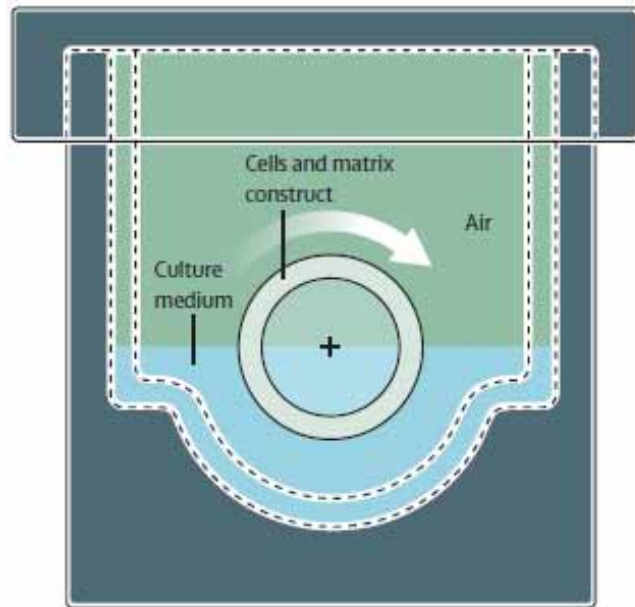
IV Graft Preparation

- Preparation of airway matrix
- Bioreactor design
- Seeding
 - Internal surface: autologous epithelial cells
 - Isolated and grown from bronchoscopic biopsy samples
 - External surface: autologous chondrocytes
 - Induced **differentiation** of mesenchymal stem cells from bone-marrow aspirate

Clinical transplantation of a tissue-engineered airway

IV Graft Preparation

- Preparation of airway matrix
- Bioreactor design
- Seeding



Clinical transplantation of a tissue-engineered airway


V Graft Implantation and Results

- Evolution was rapidly favorable
 - Improved quality of life
 - Improved Lung Function tests

	December, 2007 (before surgery)	August, 2008 (2 months after surgery)	September, 2008 (3 months after surgery)
FVC (L)	2.35 (62%)	3.81 (100%)	3.86 (100%)
FEV ₁ (L)	1.75 (55%)	3.17 (100%)	3.25 (100%)
FEV ₁ /FVC	0.74	0.95	0.99
Raw (kPa/Lxs)	5.57	3.06	3.31
SGaw (kPa ⁻¹ s ⁻¹)	0.058	0.122	0.213

A substantial reduction in forced expiratory volume in 1 s (FEV₁) and a smaller but significant reduction in forced vital capacity (FVC) occurred preoperatively. A complete reversal of airway obstruction, a substantial amelioration in airway resistance and conductance occurred postoperatively. Ratio of FEV₁ to FVC in healthy adults is 0.75–0.80; Raw=airway resistance. SGaw=specific airways conductance.


Table: Lung function test results



Clinical transplantation of a tissue-engineered airway

V Graft Implantation and Results

- Evolution was rapidly favorable
 - Improved quality of life
 - Improved Lung Function tests
 - Endoscopic monitoring of the graft
 - Normal mucosa secretion
 - No inflammation
 - Laser Doppler: rich microvascular bed



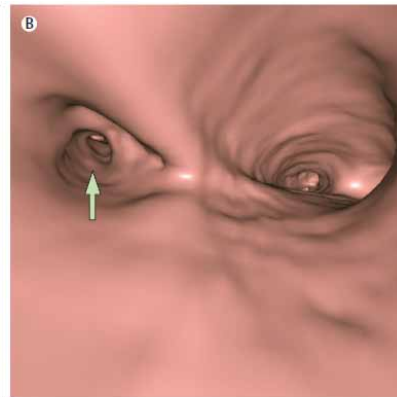
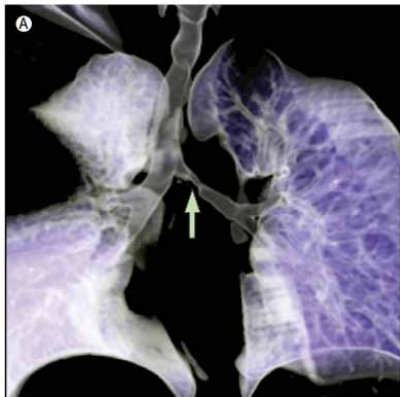
Clinical transplantation of a tissue-engineered airway

V Graft Implantation and Results

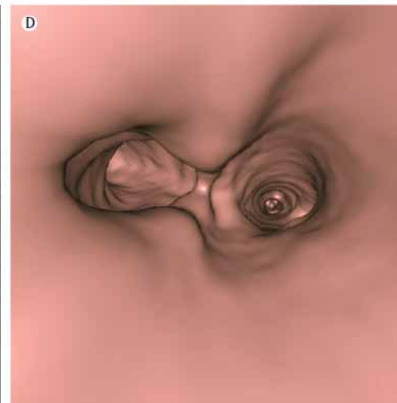
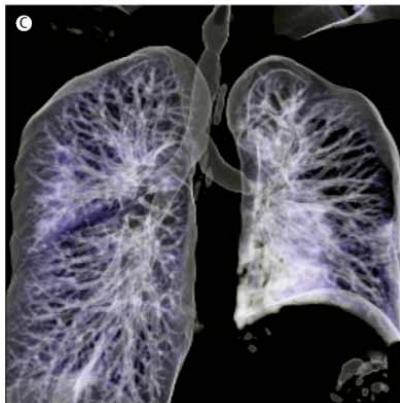
- Evolution was rapidly favorable
 - Improved quality of life
 - Improved Lung Function tests
 - Endoscopic monitoring of the graft
 - CT-reconstruction at 1 month post-op
 - Wide potency

Clinical transplantation of a tissue-engineered airway


V Graft Implantation and Results



CT-reconstruction:
stenosis




CT-reconstruction
at 1 month post-op



Clinical transplantation of a tissue-engineered airway

V Graft Implantation and Results

- Evolution was rapidly favorable
 - Improved quality of life
 - Improved Lung Function tests
 - Endoscopic monitoring of the graft
 - CT-reconstruction at 1 month post-op
 - Serology
 - Absence of anti-donor HLA antibodies at 14 days, 1 & 2 months



Clinical transplantation of a tissue-engineered airway

VI Conclusion & Perspectives

- Successful tissue-engineered large airway replacement
 - Improved quality of life & respiratory function
 - No immunosuppressive therapy needed
 - 4 month follow-up



Clinical transplantation of a tissue-engineered airway

VI Conclusion & Perspectives

- Successful tissue-engineered large airway replacement
- Clinical trial



Clinical transplantation of a tissue-engineered airway

VI Conclusion & Perspectives

- Successful tissue-engineered large airway replacement
- Clinical trial
- Feasibility of tissue engineering based on the combination of
 - Autologous cells
 - Decellularized tissue matrix